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10822300
1/10/08

Appl. No. 10/822,300
Amdt. dated May 1, 2006
Reply to Office Action of December 30, 2005

PATENT

Biosciences). Mean channel fluorescence (MCF) of each mutant was plotted using Excel (Microsoft® Corporation).

Please replace the paragraph at page 73, lines ¹⁶~~14~~-29, with the following amended paragraph:

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A dilution series of each purified OST577-IgG2M3 antibody was competed against biotinylated OST577-IgG2M3 antibody for binding to human FcRn on cell line NS0 HuFcRn (memb), clone 7-3 at 37° C. Approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 100 μ l of pre-mixed biotinylated OST577-IgG2M3 antibody (10 μ g/ml) and OST577-IgG2M3 competitor antibody (twofold serial dilutions, from 208 μ g/ml to 0.102 μ g/ml) in FBB, pH 6.0. The cells were incubated with the antibody mixture for 1 hour at 37° C., washed twice in FBB, pH 6.0, and resuspended in 25 μ l of streptavidin-RPE conjugate (BioSource International) diluted to 2.5 μ g/ml in FBB, pH 6.0. After incubation for 30 minutes in the dark, the cells were washed twice in FBB, pH 6.0, and resuspended in 1% formaldehyde. Samples were analyzed for antibody binding to FcRn by FACS™ using a FACScan™ flow cytometer (BD® Biosciences). Mean channel fluorescence (MCF) was plotted against competitor concentration, and IC50 values were calculated using GraphPad Prism® (GraphPad™ Software).

Please replace the paragraph beginning at page 73, line 31, with the following amended paragraph:

Purified OST577-IgG2M3 and OST577-IgG1 mutant antibodies were compared to the respective wild-type antibodies for binding to human FcRn and then released at various pH values in single-point binding and release assays using cell line NS0 HuFcRn (memb), clone 7-3. Approximately 2×10^5 cells/test were washed once in FBB, pH 8.0, and once in FBB, pH 6.0, then resuspended in 100 μ l of purified antibody (10 μ g/ml) in FBB, pH 6.0. The cells were